

Open Space Advisory Committee
Applicant Package - Regular Member

Open Space Advisory Committee - Regular Member

Term: 14 Mar 2022 - 25 Jan 2025

Positions Available: 3

Number of applicants in this package: 1

- Bublitz, DeAnna

Name: Bublitz, DeAnna

Mailing Address: 1102 S 5th St W, Missoula/MT, 59801

Email Address: deannabublitz@gmail.com

Board Name: Open Space Advisory Committee

Daytime Phone:: 612-790-2920

How long have you been a city resident?: 7 years

In which ward do you reside?: Ward 3

Are you registered to vote?: Yes

Current Occupation: Research Professor

Current Employer: University of Montana

Briefly describe your educational background: I have a Bachelor of Science in biology and hold a Ph.D. in microbiology and molecular genetics. I currently research infectious diseases at the University of Montana.

List of community service experience: - Founder of D.E.E.R. Camp – MT
I founded a group that works to reduce the barriers to get into hunting in Montana, especially for folks in groups historically excluded from the activity. We have led summer conservation programs with the Y.W.C.A., skills workshops with MUD and Queers & Camo, and sat on the “Voices in Conservation” panel hosted by Montana Wildlife Federation. The current focus is to create a gear library to reduce the financial barrier for folks just getting into hunting. We are partnering with Hellgate Hunters and Anglers to get this off the ground.

- Ambassador – Montana Wildlife Federation (~7 mos)
Act as a representative for Montana Wildlife Federation (MWF) in my personal conservation activities as well as MWF organized events. Write letters to the editor, be involved in legislative activities in Montana, and broadly work to expand the community of conservationists involved with MWF.

- MUD Board of Directors Vice President and member of Operations & Finance Subcommittee Board (~3 years)
Guide the management of the nonprofit MUD, that serves to educate, demonstrate, and encourage sustainable urban living through a tool loan program, workshops, and community events. Operations & Finance subcommittee oversees financial investments and infrastructure development.

- Open Space Advisory Committee – City of Missoula (~ 4 years)

- Girls Using Their Strengths (GUTS) Volunteer – Y.W.C.A. (4 years)
Lead weekly group sessions of 1 – 1.5 hours during or after school with 4th - 8th graders from area schools. Focus on giving young girls the tools to practice leadership, learn about healthy relationships, explore their own strengths and become empowered members of their communities.

What is your interest in serving on this board, commission or committee?: I have been on the open space committee for the last 4 years and volunteered for the subcommittee that helped redesign our rubric used to evaluate projects asking for open space money. I have loved working with the folks on this committee really enjoyed being a part of the city process that maintains conservation lands for the health of our environment as well as recreation areas for Missoulians. I am very interested in staying on the committee to see our new rubric in action and be a part of the team as we begin using the 2018 bond and levy dollars. I am also a regular user of our open spaces as a runner, hiker, dog walker, biker, and want to see them continue to provide quality outdoor space for Missoulians into the future.

What special knowledge, interest, or experience do you possess that would qualify you for a position on this board commission, or committee?: I have been a part of OSAC for the past 4 years and work well with the committee members and city staff. I

have attended nearly all meetings and fieldtrips in those 4 years and have further volunteered time to remake our evaluation rubric, attend trail openings, and help re-seed the Waterworks burn area this past fall. More broadly, I am invested in my Missoula community and conservation issues on the whole. As an ambassador for MWF and with my own organization I participate in several conservation and hunting-related initiatives including building a new pit toilet at Heart Lake with the Great Burn Conservation Alliance and leading GUTS summer groups in outdoor wildlife ID and tracking exercises. In all my volunteer endeavors I am striving to make Missoula more sustainable and equitable place which are key aspects of open spaces in Missoula.

Reference 1 Name: Marcus Strange (MT Wildlife Federation)

Reference 2 Name: Casey Valencia (MUD)

Attachments:

- Bublitz_CV.pdf

DeAnna Bublitz

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Education:

Ph.D. – Microbiology and Molecular Genetics - Stony Brook University, NY May 2012
B.S. – Major: Biology; Minor: Chemistry - University of Wisconsin – Stevens Point, WI Dec 2006

Summary of Qualifications:

- Ph.D. in microbiology and molecular genetics with extensive experience in identifying and implementing information and ideas from many fields in order to develop cutting edge and creative approaches to protocols, data evaluation, and accomplishing my scientific goals
 - 17 years of biological laboratory experience spanning academia, government, private, and educational labs as well as applied and curiosity-focused research
 - Worked in fields of: bacteriophages, host-pathogen interaction, endosymbiont evolution and organellogenesis, and field work to detect pathogens and mitigate transmission
 - Extensive experience extracting and analyzing DNA and RNA from: bacteria, insects, mammalian tissues, feces, filter papers, soil/sediment, and fish tissue. Analysis via: PCR, quantitative PCR, and Sanger and Illumina sequencing
 - Wide-ranging knowledge of laboratory procedures including genomic analysis, cloning, confocal microscopy, TEM, nanoSIMS, tissue culture (primary and cell lines), DNA extraction and analysis, ELISA and western blots
 - Strong interpersonal skills having provided leadership and professional expertise to teams of interdisciplinarity and international teams in India and Madagascar. Also have five years of experience leading and training students and new hires in an academic lab setting
 - Expertise in independently planning, organizing, and conducting the full cycle of research, from idea generation and study design, to implementation, documenting findings, and interpreting results, and grant acquisition. I have disseminated my work in both peer-reviewed primary and review articles, conference posters, and presentations
 - Act as principal investigator for a pilot project yielding experience in understanding of initiating research projects, the relationships of organizations in health research, and managing a budget
-

Professional Experience:

Research Assistant Professor – Promotion in same lab; Aug 2021- Current

Department of Biological Sciences, University of Montana, Missoula, MT

Dates of Employment: Aug 2021 – current (40 hrs/week)

Supervisor: Dr. Patrick Secor

Project: Investigate the function of bacteriophage and their associated retrons in eukaryotic systems for health-related research

Major Duties:

- Serve as principal investigator health and vaccine-related research for infectious diseases and their related innate immune responses
- ISE6 (tick cell line) and RAW 264.7 (mouse cell line) culture
- Conduct innate immune-response experiments, bacterial survival assays with *Borrelia burgdorferi*, *Pseudomonas aeruginosa*, and their related bacteriophages
- Generate mutant strains of *P. aeruginosa*, perform ELISAs, PCR/RT-PCR/qPCR, BrdU uptake, and various other molecular and cell biology assays
- Keep detailed records of experimental data, tabulate, summarize, and analyze data using Prism, Microsoft Excel, RStudio, and Illustrator
- Aid in manuscript and grant writing/review

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- Utilize scientific knowledge and expertise to perform peer review, train lab members in various microbiological and cell culture techniques; train and supervise a rotation student
- Obtained project grant for *B. burgdorferi* project: \$75,000; developed and manage the budget for this project

Senior Research Scientist – Department of Biological Sciences, University of Montana, Missoula, MT

Dates of Employment: Aug 2020 – Aug 2021(40 hrs/wk)

Supervisor: Dr. Patrick Secor

Project: Investigate the function of bacteriophage and their associated retrons in eukaryotic systems

Major Duties:

- ISE6 (tick cell line) and RAW 264.7 (mouse cell line) culture
- Conduct innate immune-response experiments, bacterial survival assays with pathogens *Borrelia burgdorferi*, *Pseudomonas aeruginosa*, and their related bacteriophages
- Generate mutant strains of *P. aeruginosa*, perform ELISAs, PCR/RT-PCR/qPCR, BrdU uptake, and various other molecular and cell biology assays
- Develop protocols for non-model organisms and cell types
- Keep detailed records of experimental data, tabulate, summarize, and analyze data using Prism, Microsoft Excel, RStudio, and Illustrator
- Train lab members in various microbiological and cell culture techniques; train and supervise a rotation student
- Aid in manuscript and grant writing/review

Senior Research Scientist/Lab Manager – Department of Biological Sciences, University of Montana, Missoula, MT

Dates of Employment: Aug 2018 – August 2020 (40 hrs/week)

Supervisor: Dr. John McCutcheon

Project: Investigate the evolution of bacterial endosymbionts of insects

Major Duties:

- Maintain S2 cell cultures (insect cell line), bacterial cell lines, and mealybug colonies
- Isolate DNA and RNA from insects and bacteria for transcriptome analysis and PCR
- Conduct infection assays, growth analysis, and flow cytometry analysis of S2 cells and bacterial cultures
- Process and image S2 cells, dissect and process insects for genomics and imaging on various microscopes including confocal, dissection, and transmission electron (TEM)
- Creating and establishing novel protocols to address the needs of my experiments and scientific questions in a non-model organism
- Keep detailed records of experimental data, tabulate, summarize, and analyze data using Microsoft Excel, RStudio, and Illustrator
- Manage daily lab needs such as ordering supplies, training new individuals, maintain training records, and servicing equipment; manage budget
- Collaborate with multiple institutes; communicate findings through peer-reviewed publications, conference-presented posters, and seminar talks

Postdoctoral Fellow – Department of Biological Sciences, University of Montana, Missoula, MT

Dates of Employment: May 2016 – Aug 2018 (promoted to research scientist/lab manager; 40 hrs/week)

Supervisor: Dr. John McCutcheon

Project: Investigate the evolution of bacterial endosymbionts of insects

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Major Duties:

- Maintain insect cultures, insect dissection and downstream processing for genomics and imaging on various microscopes including confocal, dissection, and TEM
- Isolate DNA and RNA from insects and bacteria for PCR, genomic library preparation and genome sequence and transcriptome analysis
- Bacterial genome library preparation, sequencing, assembly, and analysis
- Field collection and processing of cicada species in Montana and Arizona
- Creating and establishing novel protocols to address the needs of my experiments and scientific questions in a non-model organism
- Keep detailed records of experimental data, tabulate, summarize, and analyze data using Microsoft Excel, RStudio, and Illustrator
- Communicate with collaborators both on campus and elsewhere to conduct experiments and write manuscripts; communicate findings through peer-reviewed publications, conference-presented posters, and seminar talks

Biological & Genetics Technician – University of Montana/United States Forest Service, Rocky Mountain Research Station, Missoula, MT

Dates of Employment: Aug 2015 – May 2016 (20+ hrs/week)

Supervisor: Dr. Kellie Carim

Project: Environmental DNA (eDNA) analysis in aquatic systems

Major Duties:

- Processed eDNA samples as a part of ecosystem management and conservation studies being run throughout the United States focused primarily on endangered or invasive fish species
- Isolated eDNA from soil, sediment, and filter paper
- Used quantitative PCR instruments to assay extracted DNA; helped design primers for qPCR detection of various aquatic species
- Analyzed data for presence of select aquatic species
- Kept detailed records of samples, data, results, and other information using Microsoft Excel and Word
- Maintained eDNA water pump equipment, assembled sample acquisition kits for collaborators
- Aided in writing summary reports for stakeholders

Adjunct Professor – University of Montana/Bitterroot College, Missoula/Hamilton, MT

Dates of Employment: Various 2013 – 2016 (12+ hrs/week)

Supervisor: Dr. Victoria Clark

Major Duties:

- Taught full semester courses of introductory anatomy & physiology lecture, introductory chemistry lab, and introductory biology lecture and lab at the Bitterroot College
- Guest-taught introductory biology lecture at the main campus for one month
- Details for each course can be found in the Teaching section

Study Director of Vector-borne Diseases – Genesis Labs, Wellington, CO and Patna, Bihar, India

Dates of Employment: Dec 2014 – Aug 2015

Supervisor: Richard Poche

Project: Test a product meant to mitigate transmission of the infectious disease visceral leishmaniasis in India

Major Duties:

- Managed team of ten people from India, Belgium, and the United States to conduct field work, data collection, and data analysis

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- Planned and executed field studies involving human behavior surveys, serology tests, insect (sand fly) capture, and treating cattle in three villages in remote regions of northern India
- Collect data concerning sand fly populations including collection counts, location data, animal surveys, emergence traps, and survey for previous mitigation efforts by government and local officials
- Developed a phone application used to collect survey information (ODK/Android platform), created user manual and trained team how to use the app in the field
- Isolated DNA from insects, developed primers, and conducted PCR and electrophoresis for blood-meal analysis, species identification, and presence of *Leishmania donovani* – conducted related data analysis
- Kept detailed records of data, contracts, and meetings
- Acted as liaison between various Indian government, nonprofit organizations, and Genesis Labs
- Wrote and published one review article and aided in grant writing

Postdoctoral Fellow – Rocky Mountain Laboratories (RML), National Institutes of Allergy and Infectious Disease/National Institutes of Health, Hamilton, MT

Dates of Employment: Aug 2012 – Nov 2014 (40 hrs/week)

Supervisor: Dr. Ted Hackstadt

Project: Explore the virulence mechanisms of the pathogen *Rickettsia rickettsii*

Major Duties:

- Planned and executed experiments in Biosafety level 2 and level 3 labs.
- Cultured and maintained primary endothelial cells; performed infection assays and related analyses with *Rickettsia rickettsii*
- Extracted DNA and RNA from *R. rickettsii* and associated infected cells for microarray plate analysis, cloning proteins of interest, and sequencing genes of interest
- Cloned genes of interest into expression plasmids for protein expression and western blot analysis
- ELISA of conditioned media from infected cells to detect various immune response markers
- Constructed controlled expression vectors for genes of interest, imaged infected cells by confocal microscope
- Additional work involving: flow cytometry, mammalian tissue culture, and animal infection assays
- Assisted in comparative genome analysis of *Rickettsia rickettsii* strains
- Wrote and published a peer-review article, presented work at department seminars

Independent Researcher – Emory University, Atlanta, GA; Stony Brook University, Stony Brook, NY; Centre ValBio, Ranomafana, Madagascar

Dates of Employment: Jun & Jul 2011 and 2012 (Field work in 2011, DNA analysis and writing 2012; 40 hrs/week)

Supervisors: Drs. James Bliska and Thomas Gillespie

Project: Investigate transmission of infectious diseases between people, the environment, and domestic and wild animals

Major Duties:

- Worked with a team of disease ecologists, health and hygiene workers, and research technicians in three villages and Ranomafana National Park in Madagascar
- Developed research proposal, budget and acquired funding from Stony Brook Hospital (\$1000) to assess bacterial contamination of water sources
- Isolated and analyzed DNA from fecal samples by PCR and gel electrophoresis for presence of enteric pathogens

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- Worked in a team conducting household interviews, small mammal trapping/handling, blood collection from small mammals, and collection and processing of water and fecal samples
- Helped develop, code, and analyze surveys for human subjects; acquired IRB approval from Stony Brook University; trained technicians to administer survey in the field
- Wrote and published two peer-reviewed articles based on results

Research Assistant (Doctoral Work) – Stony Brook University, Stony Brook, NY

Dates of Employment: Aug 2007 – May 2012 (40 hrs/week)

Supervisor: Dr. Martha Furie

Project: Study *Francisella tularensis* (causative agent of tularemia) manipulates the human innate immune

Major Duties:

- Isolate and culture primary human endothelial and macrophage cells
- Planned and executed experiments using immunology and cell-biology techniques: mammalian tissue culture, flow cytometry, ELISA, microarray analysis, DNA and RNA isolation, qRT-PCR, PCR, gel electrophoresis, cloning, western blot, confocal microscopy, data analysis
- Identified bacterial mutants of interest, performed infection and immunoassays as well as growth curves
- Collaborated with outside labs to express proteins of interest, acquire knock-out strains of mice
- Performed DNA extraction on mouse tail snips and PCR for validation of knock-out status
- Mentored first year graduate students on proper lab protocol, assay techniques, and project design
- Kept detailed records of data; analyzed and tabulated using Microsoft Excel, Photoshop and SigmaPlot
- Conducted infection assays and related processing and analyses in BSL2 and BSL3 laboratories
- Wrote and published three peer-reviewed articles based on results
- Teaching assistant for three semesters of microbiology labs for nursing and medical students

Molecular Research Assistant – University of WI – Stevens Point, WI

Dates of Employment: Aug 2005 – Dec 2006 (8 hrs/week)

Supervisor: Dr. Diane Caporale

Project: Study the evolution of *Borrelia burgdorferi* in Wisconsin via analysis of the *ospB* gene

- Isolated *Borrelia burgdorferi* (causative agent of Lyme disease) DNA from ticks from various locations in Wisconsin
- Performed inoculation, data collection, dissection, embedding, and histologic slide preparation for a mouse model experiment to test pathogenicity of a variant strain
- Sequenced and analyzed genes of *B. burgdorferi* to characterize a variant strain and determine strain variation across Wisconsin. Data were used to determine tick population structure and mobility

Lab and Data Analysis Experience:

Lab experience: Proper sterile technique and handling for DNA and RNA extraction from many tissue types including: bacteria, insects, mammalian tissues, cell cultures, feces, filter papers, soil/sediment, and fish tissue; primer design for PCR and qPCR; DNA plasmid prep, cloning, transformation, and sequencing; insect mitochondrial and bacterial genome analysis; bacterial genome assembly from DNA extraction, library preparation, to Sanger and Illumina sequence preparation including: Qbit, Blue Pippin, spectrophotometers and plate readers, thermocyclers; related genome assembly, genetic and

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phylogenetic bioinformatic analysis; extensive tissue culture experience using primary human endothelial and macrophage cells and cell lines (HeLa, S2, J774); Protein-based methods including ELISA, protein expression, and western blot; understanding of, and ability to apply, general microbiological and cellular biological methods. Infectious disease assays, protocol development, grant proposal, management, and budget development/management.

Analysis and software experience: Genetic and genomic sequence assembly and analysis using: MEGA, Geneious and CodonCode; proficient using BLAST, both NCBI user interface and via Terminal; Illumina read processing and analysis using Terminal packages: Trimmomatic, SPAdes, FastQC, Promer/Mummerplot, samtools, PEAR, and Tablet; phylogenetic tree assembly via mafft and RAxML; basic data analysis and visualization (t-tests, jitter plots) with R and ggplot. Strong Illustrator, Photoshop, and Microsoft Word, Excel, and PowerPoint skills.

Peer-Reviewed Publications:

Secor, P.R., Michaels, L.A., **Bublitz, D.C.**, Jennings, L.K., and Singh, P.K. (*Preprint*). The depletion mechanism can actuate bacterial aggregation by self-produced exopolysaccharides and determine species distribution and composition in bacterial aggregates. *bioRxiv* 2021.05.11.443568; doi: <https://doi.org/10.1101/2021.05.11.443568>.

Macias, A.M., Geiser, D.M., Stajich, J.E., Łukasik, P., Veloso, C., **Bublitz, D.C.**, Berger, M.C., Boyce, G.R., Hodge, K., Kasson, M.T. (2020). Evolutionary relationships among *Massospora* spp. (Entomophthorales), obligate pathogens of cicadas. *Mycologia*. 112: 1060-1074.

Bublitz D.C., Chadwick, G.L., Magyar, J.S., Sandoz, K.M., Brooks, D.M., Mesnage, S., Ladinsky, M.S., Garber, A.I., Bjorkman, P.J., Orphan, V.J., and McCutcheon, J.P. (2019) Peptidoglycan production by an insect-bacterial mosaic. *Cell*. 179: 703-712.

Łukasik, P., Chong, R.A., Nazario, K., Matsuura, Y., **Bublitz, D.C.**, Campbell, M.A., Meyer, M.C., Van Leuven, J.T., Pessacq, P., Veloso, C., Simon, C., and McCutcheon, J.P. (2019) One Hundred Mitochondrial Genomes of Cicadas, *Journal of Heredity*. 110: 247–256.

Bublitz DC, Poché RM, and Garlapati R. (2016). Measures to control *Phlebotomus argentipes* and visceral leishmaniasis in India. *Journal of Arthropod-Borne Diseases*. 10: 113-126.

Clark, T. C., Noriea, N. F., **Bublitz, D. C.**, Ellison, D. W., Martens, C., Lutter, E. I., and Hackstadt, T. (2015). Comparative genome sequencing of *Rickettsia rickettsii* strains that differ in virulence. *Infection and Immunity*. 83: 1568-1576.

Bublitz, D. C., P. C. Wright, F. T. Rasambainarivo, S. J. Arrigo-Nelson, J. R. Bodager, and T. R. Gillespie (2014) Pathogenic enterobacteria in lemurs associated with anthropogenic disturbance. *American Journal of Primatology*. DOI: 10.1002/ajp.22348.

Bublitz, D. C., P. C. Wright, J. R. Bodager, F. T. Rasambainarivo, J. B. Bliska, and T. R. Gillespie (2014) Epidemiology of pathogenic enterobacteria in humans, livestock, and peridomestic rodents in rural Madagascar. *PLoS One*. 9: e101456.

Bublitz, D. C., C. E. Noah, J. L. Benach, and M. B. Furie (2010) *Francisella tularensis* suppresses the proinflammatory response of endothelial cells via the endothelial protein C receptor. *Journal of Immunology*. 185: 1124-1131.

Platz, G.J., **Bublitz, D. C.**, Mena, P., Benach, J.L., Furie, M.B., and Thanassi, D.G. (2010) A tolC mutant of *Francisella tularensis* is hypercytotoxic and elicits increased proinflammatory responses from host cells. *Infection and Immunity*. 78: 1022-1031.

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Noah, C.E., Malik, M., **Bublitz, D. C.**, Camenares, D., Sellati, T.J., Benach, J.L., and Furie, M.B. (2010) GroEL and lipopolysaccharide from *Francisella tularensis* live vaccine strain synergistically activate human macrophages. *Infection and Immunity*. 78: 1797-1806.

Popular Press Articles:

Callier, V. Cell-Bacteria Mergers Offer Clues to How Organelles Evolved. Quanta Magazine. 03 October 2019. <https://www.quantamagazine.org/cell-bacteria-mergers-offer-clues-to-how-organelles-evolved-20191003>.

Williams, R. When Is an Endosymbiont an Organelle? The Scientist. 03 October 2019. <https://www.the-scientist.com/news-opinion/endosymbiont-or-organelle--66526>.

Radkov A. D. and Chou, S. (2019) An Affair to Remember: How an Endosymbiont Partners with Its Host to Build a Cell Envelope. Cell. 179: 584-586.

Paniagua, T. Summer's Insects Draw Scientists to Southern Arizona. 28 August 2017. <https://news.azpm.org/s/50274-summers-insects-draw-scientists-to-southern-arizona/>.

Teaching:

Principles of Living Systems (BIO 160) – University of Montana – main campus, MT Oct 2016

- Prepare and present lectures on the fundamentals of biology for ~400 biology-major undergraduate students
- Devise and administer exam
- One month of a full semester course to cover for a professor on a field study

Discover Biology (BIO 101) – University of Montana – Bitterroot College, MT 2015-2016

- Prepare and present lectures on the fundamentals of biology for ~20 biology non-major undergraduate and certificate students
- Create and lead laboratory activities based on the lecture concepts
- Devise and administer exams
- Full semester course – lab and lecture; taught two semesters

Basic Anatomy & Physiology (BIO 108) – University of Montana – Bitterroot College, MT 2013-2014

- Prepare and present lectures on anatomy and physiology for ~20 undergraduate/certificate students
- Devise and administer exams
- Full semester course; taught two semesters

General Chemistry (CHMY 122) – University of Montana – Bitterroot College, MT 2013-2014

- Prepare and present brief lab-related lectures on general chemistry concepts for ~12 undergraduate/certificate students
- Create and lead laboratory activities based on the lecture concepts
- Devise and administer exams, full semester course; taught two semesters

Biomedical Research After School Scholars (BRASS) - RML, NIAID/NIH, MT 2012- 2014

- Prepared and presented lectures and lab activities pertaining to environmental biology for groups of children grades 6-8 at three middle schools in Ravalli County, Montana

Medical Microbiology Lab (HBM 531) – Stony Brook University, Stony Brook, NY 2008, 2011

- Explained and demonstrated clinical microbiological procedures for ~80 medical and dental students
- Set up and took down lab gear, graded lab reports;

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- Two-week course

Microbiology Lecture (HBM 320) – Stony Brook University, Stony Brook, NY 2008

- Proctored exams for upper level, undergraduate microbiology lecture class
- Full semester course

Microbiology Lab (HBM 320) – Stony Brook University, Stony Brook, NY 2008

- Responsible for teaching ~40 students in undergraduate microbiology lab for pre-nursing program
- Set up and took down lab gear, explained and demonstrated basic microbiological procedures, wrote weekly quizzes, graded assigned lab reports and exams
- Full semester course

Genetics Tutor – University of WI – Stevens Point, Stevens Point, WI 2006

- Led group and one-on-one tutoring sessions for undergraduate genetics course
-

Selected Presentations:

TALKS

Peptidoglycan and the evolution of eukaryotic organelles. NASA Astrobiology Institute Team Meeting. Georgia Institute of Technology. January 2020. Atlanta, GA.

Peptidoglycan and the evolution of eukaryotic organelles. CHALK/AEGG Seminar Series – Organismal Ecology and Evolution Dept. University of Montana. October 2019. Missoula, MT.

POSTERS

Publitz, D. C., Peptidoglycan is made by an insect-bacterial mosaic. Great Wall Symposium. September 2019. Paris, France.

Publitz, D. C., The tip of the iceberg: lineage splitting in *Hodgkinia*. International Symbiosis Society Congress. July 2018. Corvallis, OR.

Publitz, D. C., Independent lineage splits in the cicada endosymbiont *Hodgkinia cicadicola* - a common occurrence. Animal-Microbe Symbioses Gordon Research Conference. June 2017. West Dover, VT.

Publitz, D. C. *Francisella tularensis* inhibits the proinflammatory activation of endothelial cells. Microbial Adhesion and Signal Transduction Gordon Conference. July 2011. Newport, RI.

Publitz, D. C. How *Francisella tularensis* – an infectious bacterium – inhibits our immune response. Research that Matters Exposition. March 2011. Albany, NY

Publitz, D. C. *Francisella tularensis* inhibits the proinflammatory activation of endothelial cells via the endothelial protein C receptor. 110th General Meeting of the American Society for Microbiology. May 2010. San Diego, CA.

Publitz, D. C. *Francisella tularensis* inhibits the proinflammatory activation of endothelial cells via the endothelial protein C receptor. 6th International Conference on Tularemia. September 2009. Berlin, Germany.

Current Volunteer Activities:

Founder of D.E.E.R. Camp – MT – Missoula, MT

Sept 2020 – Current

I founded a group that works to reduce the barriers to get into hunting in Montana, especially for folks in groups historically excluded from the activity. We have led summer conservation programs with the Y.W.C.A., skills workshops with MUD and Queers & Camo, and sat on the “Voices in Conservation” panel hosted by Montana Wildlife Federation. The current focus is to create a gear library to reduce the financial barrier for folks just getting into hunting.

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Ambassador – Montana Wildlife Federation, Helena, MT Aug 2021 – Current
Act as a representative for Montana Wildlife Federation (MWF) in my personal conservation activities as well as MWF organized events. Write letters to the editor, be involved in legislative activities in Montana, and broadly work to expand the community of conservationists involved with MWF.

MUD Board of Directors – Missoula Urban Demonstration, Missoula, MT Mar 2020 – Current
Board Vice President and member of Operations & Finance Subcommittee
10-member board that guides the management of the nonprofit MUD, that serves to educate, demonstrate, and encourage sustainable urban living through a tool loan program, workshops, and community events. Operations & Finance subcommittee oversees financial investments and infrastructure development.

Open Space Advisory Committee – City of Missoula, Missoula, MT Feb 2018 – Current
12-member committee that helps implement the Missoula Urban Area Open Space Plan. Evaluate and provide recommendations for land acquisitions and trail development and improvement concerning open space conservation. Elected vice chair for subcommittee to overhaul project evaluation criteria.

Girls Using Their Strengths (GUTS) Volunteer – Y.W.C.A., Missoula, MT Sept 2016 – 2020
Lead weekly group sessions of 1 – 1.5 hours during or after school with 4th - 8th graders from area schools. Focus on giving young girls the tools to practice leadership, learn about healthy relationships, explore their own strengths and become empowered members of their communities.
